

**ЦИТИРАНИЯ**  
 (без автоцитати и полуцитати)  
**гл. ас. д-р Гергана Кирилова Михайлова**

Справката е генерирана от SONIX, системата за отчитане на научната и експертната дейност в БАН

**E 1.8.1:**

**Цитати (първа част - на научни публикации) - в WoS или Scopus**

- **Звено:** ( ИФРГ ) Институт по физиология на растенията и генетика
- **Секция:** ( ИФРГ ) Фотосинтеза – активност и регулация
- **Име:** ( ИФРГ/0066 ) Михайлова, Гергана
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- **Година:** 2006 ÷ 2024
- **Тип записи:** Всички записи

Брой цитирани публикации: 32	Брой цитиращи източници: 196	Коригиран брой: 196.000
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**2008**

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1. Péli, E, Mihailova, G, Petkova, S, Georgieva, K. Root respiration in whole Haberlea rhodopensis Friv. plants during desiccation and rehydration. Acta Biologica Szegediensis, 52, 1, University of Szeged, 2008, ISSN:1588-385X, SCOPUS, 115-117

Цитира се в:

1. Baloutzov V, Gemishev T, Tsvetkov T. A study of the composition of the biologically active substances in Haberlea rhodopensis Friv, Comptes Rendus de L'Academie Bulgare des Sciences, 62, 5, 585-588, @2009 [Линк](#)
2. Zheleva-Dimitrova, D., Nedialkov, P., & Giresser, U. (2016). A Validated HPLC Method for Simultaneous Determination of Caffeoyl Phenylethanoid Glucosides and Flavone 8-C-glycosides in Haberlea rhodopensis. Natural product communications, 11(6), 1934578X1601100622., @2016 [Линк](#)

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**2009**

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2. Mihailova, G, Petkova, S, Georgieva, K. Changes in some antioxidant enzyme activities in Haberlea rhodopensis during desiccation at high temperature. Biotechnology & Biotechnological Equipment, 23, 1, Taylor & Francis, 2009, ISSN:1310-2818, Web of Science, DOI:10.1080/13102818.2009.10818487, 561-564. ISI IF:0.3

Цитира се в:

3. Djiljanov D., Ivanov S., Moyankova D., Miteva L., Kirova E., Alexieva V., Joudi M., Peshev D., Van den Ende W. 1.000 - Sugar ratios, glutathione redox status and phenols in the resurrection species Haberlea rhodopensis and the closely related non-resurrection species Chirita eberhardtii. Plant Biology, 13 (5), 767-776, @2011 [Линк](#)
4. Popov B., Georgieva Sv., Gadjeva V., Petrov V. - Radioprotective, anticlastogenic and antioxidant effects of total extract of Haberlea Rhodopensis on rabbit blood samples exposed to gamma radiation in vitro. Revue de Médecin Vétérinaire, 162 (1), 34-39, @2011 [Линк](#)

5. Ma HL, Xu XH, Zhao XY, Liu HJ, Chen H. Impacts of drought stress on soluble carbohydrates and respiratory enzymes in fruit body of Auricularia auricular, Biotechnology & Biotechnological Equipment, 29, 1, 10-14, @2015 [Линк](#)
6. Staneva, D., Dimitrova, N., Popov, B., Alexandrova, A., Georgieva, M., & Miloshev, G. (2023). Haberlea rhodopensis Extract Tunes the Cellular Response to Stress by Modulating DNA Damage, Redox Components, and Gene Expression. International Journal of Molecular Sciences, 24(21), 15964., @2023 [Линк](#)

## 2011

3. **Mihailova, G**, Petkova, S, Büchel, Claudia, **Georgieva, K**. Desiccation of the resurrection plant Haberlea rhodopensis at high temperature. Photosynthesis Research, 101, 1, Springer, 2011, ISSN:0166-8595, Web of Science, DOI:10.1007/s11120-011-9644-2, 5-13. ISI IF:3.502

Цитира се 6:

7. Daskalova E., Dontcheva S., Yahoubian, Minkov I., Toneva V. – A strategy for conservation and investigation of the protected resurrection plant Haberlea rhodopensis Friv. BioRisk, 6, 41-60, 2011, @2011 [Линк](#)
8. García-Plazaola J.I., Esteban R., Fernández-Marín B., Kranner I., Porcar-Castell A. (2012) Thermal energy dissipation and xanthophyll cycles beyond the Arabidopsis model, Photosynthesis Research, 113 (1-3) , 89-103, @2012 [Линк](#)
9. Gashi B, Babani F, Kongjika E. Chlorophyll fluorescence imaging of photosynthetic activity and pigment contents of the resurrection plants Ramonda serbica and Ramonda nathaliae during dehydration and rehydration. Physiology and Molecular Biology of Plants. 2013. 19(3), 333-341., @2013 [Линк](#)
10. Mitra J, Xu G, Wang B, Li M, Deng X. Understanding desiccation tolerance using the resurrection plant Boea hygrometrica as a model system. Frontiers in plant science. 2013. doi: 10.3389/fpls.2013.00446, @2013 [Линк](#)
11. Singh P, A Tiwari, SP Singh, RK Asthana (2013) Desiccation induced changes in osmolytes production and the antioxidative defence in the cyanobacterium Anabaena sp. PCC 7120. Physiology and Molecular Biology of Plants, DOI 10.1007/s12298-012-0145-3, @2013 [Линк](#)
12. Yan K, Chen P, Shao H, Shao C, Zhao S, Breistic M. Dissection of Photosynthetic Electron Transport Process in Sweet Sorghum under Heat Stress. PLoS ONE. 2013. 8(5), e62100. doi:10.1371/journal.pone.0062100, @2013 [Линк](#)
13. Yan K, Chen P, Shao H, Shao C, Zhao S. Characterization of photosynthetic electron transport chain in bioenergy crop Jerusalem artichoke (*Helianthus tuberosus L.*) under heat stress for sustainable cultivation. Industrial Crops and Products. 2013. 50, 809-815. http://dx.doi.org/10.1016/j.indcrop.2013.08.012., @2013 [Линк](#)
14. Li, X., Yang, Y., Ma, L., Sun, X., Yang, S., Kong, X., ... & Yang, Y. (2014). Comparative Proteomics Analyses of Kobresia pygmaea Adaptation to Environment along an Elevational Gradient on the Central Tibetan Plateau. PLoS one, 9(6), e98410., @2014 [Линк](#)
15. Li, X., Yang, Y., Sun, X., Lin, H., Chen, J., Ren, J., ... & Yang, Y. (2014). Comparative Physiological and Proteomic Analyses of Poplar (*Populus yunnanensis*) Plantlets Exposed to High Temperature and Drought. PloS one, 9(9), e107605., @2014 [Линк](#)
16. Liu, M., Zhang, Z., Gao, H., Yang, C., Fan, X., & Cheng, D. (2014). Effect of leaf dehydration duration and dehydration degree on PSII photochemical activity of papaya leaves. Plant Physiology and Biochemistry, 82, 85-88., @2014 [Линк](#)
17. Li X, Zhou Y, Yang Y, Yang S, Sun X, Yang Y. Physiological and Proteomics Analyses Reveal the Mechanism of *Eichhornia crassipes* Tolerance to High-Concentration Cadmium Stress Compared with *Pistia stratiotes*, PLoS ONE, 10, 4, e0124304, @2015 [Линк](#)
18. Li, H., Ahammed, G. J., Zhou, G., Xia, X., Zhou, J., Shi, K., ... & Zhou, Y. (2016). Unraveling Main Limiting Sites of Photosynthesis under Below-and Above-Ground Heat Stress in Cucumber and the Alleviatory Role of Luffa Rootstock. Frontiers in Plant Science, 7, 746., @2016 [Линк](#)
19. Ivanova, Z., Sablok, G., Daskalova, E., Zahmanova, G., Apostolova, E., Yahubyan, G., & Baev, V. (2017). Chloroplast Genome Analysis of Resurrection Tertiary Relict Haberlea rhodopensis Highlights Genes Important for Desiccation Stress Response. Frontiers in plant science, 8:204, @2017 [Линк](#)

20. Verhoeven, A., García-Plazaola, J. I., & Fernández-Marín, B. (2018). Shared mechanisms of photoprotection in 1.000 photosynthetic organisms tolerant to desiccation or to low temperature. *Environmental and Experimental Botany*, 154, 66-79., @2018 [Линк](#)
21. Gashi, B., Kongjika, E., Osmani, M., & Luma, V. (2019). Activity of δ-aminolevulinic acid dehydratase at *Ramonda nathaliae* and *Ramonda serbica* plants during dehydration and rehydration. *Biología Futura*, 9(2), 1-8., @2019 [Линк](#)
22. Yu, R.-P., Wang, G.-X., Li, F., (...), Yang, C.-M., Qu, Y.-H. (2019) Photosynthesis response of resurrection plant 1.000 *Selaginella pulvinata* detached leaves to dehydration and rehydration. *Zhiwu Shengli Xuebao/Plant Physiology Journal*, 55(12), pp. 1806-1816, @2019 [Линк](#)
23. Georgiev, Y. N., Ognyanov, M. H., & Denev, P. N. (2020). The ancient Thracian endemic plant *Haberlea rhodopensis* Friv. And related species: A review. *Journal of Ethnopharmacology*, 249, 112359., @2020 [Линк](#)
24. Płażek, A., Hura, K., Hura, T., Słomka, A., Hornyák, M., & Sychta, K. (2020). Synthesis of heat-shock proteins 1.000 HSP-70 and HSP-90 in flowers of common buckwheat (*Fagopyrum esculentum*) under thermal stress. *Crop and Pasture Science*, 71(8), 760-767., @2020 [Линк](#)
25. Liu, X. W., Wang, Y. H., & Shen, S. K. (2022). Transcriptomic and metabolomic analyses reveal the altitude 1.000 adaptability and evolution of different-colored flowers in alpine *Rhododendron* species. *Tree Physiology*, 42(5), 1100-1113., @2022 [Линк](#)

**2012**

4. Georgieva, K., Doncheva, S., Mihailova, G., Petkova, S. Response of sun-and shade-adapted plants of *Haberlea rhodopensis* to desiccation. *Plant Growth Regulation*, 67, 2, Springer, 2012, ISSN:0167-6903, Web of Sciences, DOI:10.1007/s10725-012-9669-3, 121-132. ISI IF:1.672

*Цитира се е:*

26. Aasamaa, K., & Aphalo, P. J. (2016). Effect of vegetational shade and its components on stomatal responses to 1.000 red, blue and green light in two deciduous tree species with different shade tolerance. *Environmental and Experimental Botany*, 121, 94-101., @2016 [Линк](#)
27. Karbaschi MR, Williams B, Taji A, & Mundree SG. *Tripogon loliiformis* elicits a rapid physiological and structural 1.000 response to dehydration for desiccation tolerance. *Functional Plant Biology*. 43, 7, 643-655, @2016 [Линк](#)
28. Aidar, S. D. T., Chaves, A. R. D. M., Júnior, P. I. F., Oliveira, M. D. S., da Costa Neto, B. P., Junior, T. C., & 1.000 Morgante, C. V. (2017). Vegetative desiccation tolerance of *Tripogon spicatus* (Poaceae) from the tropical semiarid region of northeastern Brazil. *Functional Plant Biology*, 44(11), 1124-1133., @2017 [Линк](#)
29. John SP, Hasenstein KH. The role of peltate scales in desiccation tolerance of *Pleopeltis polypodioides*. *Planta*, 1.000 245, 1, 207-220., @2017 [Линк](#)
30. Bayçu, G., Moustaka, J., Gevrek, N., & Moustakas, M. (2018). Chlorophyll Fluorescence Imaging Analysis for 1.000 Elucidating the Mechanism of Photosystem II Acclimation to Cadmium Exposure in the Hyperaccumulating Plant *Noccaea caerulescens*. *Materials*, 11(12), 2580., @2018 [Линк](#)
31. Moustaka, J., Ouzounidou, G., Sperdouli, I., & Moustakas, M. (2018). Photosystem II is more sensitive than 1.000 photosystem I to Al3+ induced phytotoxicity. *Materials*, 11(9), 1772., @2018 [Линк](#)
32. Moustakas, M., Bayçu, G., Gevrek, N., Moustaka, J., Csatári, I., & Rognes, S. E. (2019). Spatiotemporal 1.000 heterogeneity of photosystem II function during acclimation to zinc exposure and mineral nutrition changes in the hyperaccumulator *Noccaea caerulescens*. *Environmental Science and Pollution Research*, 26(7), 6613-6624., @2019 [Линк](#)
33. Sebastiani, F., Torre, S., Gori, A., Brunetti, C., Centritto, M., Ferrini, F., & Tattini, M. (2019). Dissecting Adaptation 1.000 Mechanisms to Contrasting Solar Irradiance in the Mediterranean Shrub *Cistus incanus*. *International journal of molecular sciences*, 20(14), 3599., @2019 [Линк](#)
34. Vassileva, V., Moyanova, D., Dimitrova, A., Mladenov, P., & Djilianov, D. (2019). Assessment of leaf 1.000 micromorphology after full desiccation of resurrection plants. *Plant Biosystems-An International Journal Dealing with all Aspects of Plant Biology*, 153(1), 108-117., @2019 [Линк](#)
35. Traykova, B. D., & Stanilova, M. I. (2020). Soilless Propagation of *Haberlea rhodopensis* Friv. Using Different 1.000 Hydroponic Systems and Substrata. *Ecologia Balkanica*, 12(1), 111-121, @2020 [Линк](#)

36. Huang, H. X., Cao, Y., Xin, K. J., Liang, R. H., Chen, Y. T., & Qi, J. J. (2022). Morphological and physiological changes in *Artemisia selengensis* under drought and after rehydration recovery. *Frontiers in Plant Science*, 13, 851942., @2022 [Линк](#)
37. Fernández-Marín, B., Nadal, M., Perera-Castro, A. V., & García-Plazaola, J. I. (2024). Photosynthetic Strategies of Desiccation-Tolerant Organisms from a Functional Perspective. In *Handbook of Photosynthesis* (pp. 659-675). CRC Press., @2024 [Линк](#)
38. Xu, Y., Chen, S., Zhao, S., Song, J., Sun, J., Cui, N., ... & Qu, B. (2024). Effects of light intensity on the photosynthetic characteristics of *Hosta* genotypes differing in the glaucousness of leaf surface. *Scientia Horticulturae*, 327, 112834., @2024 [Линк](#)
5. Péli, E, **Mihailova, G**, Petkova, S, Tuba, Z, **Georgieva, K**. Differences in physiological adaptation of *Haberlea rhodopensis* Friv. leaves and roots during dehydration-rehydration cycle. *Acta Physiologiae Plantarum*, 34, 3, Springer-Verlag, 2012, ISSN:1861-1664, Web of Science, DOI:10.1007/s11738-011-0891-9, 947-955. ISI IF:1.584

Цитира се е:

39. Gashi B, Babani F, Kongjika E. Chlorophyll fluorescence imaging of photosynthetic activity and pigment contents of the resurrection plants *Ramonda serbica* and *Ramonda nathaliae* during dehydration and rehydration, *Physiology and Molecular Biology of Plants*, 19, 3, 333-341, @2013 [Линк](#)
40. Aidar SDT, Meirelles ST, Oliveira RFD, Chaves ADM, Fernandes-Júnior PI. Photosynthetic response of *poikilochlorophyllous* desiccation-tolerant *Pleurostigma purpurea* (*Velloziaceae*) to dehydration and rehydration, *Photosynthetica*, 52, 1, 124-133, @2014 [Линк](#)
41. Li A, Wang D, Yu B, Yu X, Li W. Maintenance or Collapse: Responses of Extraplastidic Membrane Lipid Composition to Desiccation in the Resurrection Plant *Paraisometrum mileense*, *PLoS ONE*, 9, 7, e103430, @2014 [Линк](#)
42. Vieira, E. A., da Cruz Centeno, D., Freschi, L., da Silva, E. A., & Braga, M. R. (2017). The dual strategy of the bromeliad *Pitcairnia burchellii* Mez to cope with desiccation. *Environmental and Experimental Botany*, 143, 135-148., @2017 [Линк](#)
43. Vieira, E. A., Silva, K. R., Oriani, A., Moro, C. F., & Braga, M. R. (2017). Mechanisms of desiccation tolerance in the bromeliad *Pitcairnia burchellii* Mez: Biochemical adjustments and structural changes. *Plant Physiology and Biochemistry*, 121, 21-30, @2017 [Линк](#)
44. Gashi, B., Kongjika, E., Osmani, M., & Luma, V. (2019). Activity of δ-aminolevulinic acid dehydratase at *Ramonda nathaliae* and *Ramonda serbica* plants during dehydration and rehydration. *Biologia Futura*, 9(2), 1-8., @2019 [Линк](#)
45. Tebele S.M., Marks R.A., Farrant J.M. (2021) Two Decades of Desiccation Biology: A Systematic Review of the Best Studied Angiosperm Resurrection Plants. *Plants*, 10(12), 2784., @2021 [Линк](#)
46. Aidar, S. D. T., Chaves, A. R. D. M., Fernandes-Júnior, P. I., & Morgante, C. V. (2023). Physiological and biochemical changes in desiccation and rehydration cycles of *Selaginella convoluta* (*Selaginellaceae*). *Rodriguésia*, 74, e00672023., @2023 [Линк](#)

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2013

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6. Velitchkova, M, Dolchinkova, V, Lazarova, D, **Mihailova, G**, **Doncheva, S**, **Georgieva, K**. Effect of high temperature on dehydration-induced alterations in photosynthetic characteristics of the resurrection plant *Haberlea rhodopensis*. *Photosynthetica*, 51, 4, Springer Netherlands, 2013, DOI:10.1007/s11099-013-0063-9, 630-640. ISI IF:1.409

Цитира се е:

47. Sugiyama VF, Silva EA, Meirelles ST, Centeno DC, Braga MR. Leaf metabolite profile of the Brazilian resurrection plant *Barbacenia purpurea* Hook. (*Velloziaceae*) shows two time-dependent responses during desiccation and recovering, *Frontiers in plant science*, 5., @2014 [Линк](#)
48. Fernández-Marín, B., Nadal, M., Gago, J., Fernie, A. R., López-Pozo, M., Artetxe, U., ... & Verhoeven, A. (2020). Born to revive: molecular and physiological mechanisms of double tolerance in a paleotropical and resurrection plant. *New Phytologist*, 226(3), 741-759., @2020 [Линк](#)

49. Xu, M., Wang, Q., Yang, F., Zhang, T., Zhu, X., Cheng, C., & Wang, H. (2022). The responses of photosynthetic light response parameters to temperature among different seasons in a coniferous plantation of subtropical China. *Ecological Indicators*, 145, 109595., [@2022](#) [Линк](#)
7. Georgieva, K., Doncheva, S., Mihailova, G., Petkova, S. Effect of light on the photosynthetic activity during desiccation of the resurrection plant *Haberlea rhodopensis*. *Photosynthesis Research for Food, Fuel and the Future*, Springer Berlin Heidelberg, 2013, ISBN:978-3-642-32033-0, DOI:10.1007/978-3-642-32034-7\_113, 536-539
- Цитира се е:
50. Kalaiarasi, V., Almeida, R. S., Coutinho, H. D., & Johnson, M. (2022). Biochemical profile of resurrection fern allies: *Selaginella wightii* Hieron and *Selaginella involvens* (SW) Spring. *Vegetos*, 1-10., [@2022](#) [Линк](#)

**2014**

8. Solti, A., Lenk, S., Mihailova, G., Mayer, P., Barócsi, A., Georgieva, K. Effects of habitat light conditions on the excitation quenching pathways in desiccating *Haberlea rhodopensis* leaves: an Intelligent FluoroSensor study. *Journal of Photochemistry and Photobiology B: Biology*, 130, Elsevier, 2014, DOI:10.1016/j.jphotobiol.2013.11.016, 217-225. SJR (Scopus):0.721, JCR-IF (Web of Science):2.96
- Цитира се е:
51. Tan, T., Sun, Y., Luo, S., Zhang, C., Zhou, H., Lin, H. Efficient modulation of photosynthetic apparatus confers desiccation tolerance in the resurrection plant *Boea hygrometrica*. *Plant and Cell Physiology*, 58(11), 1976-1990, 2017., [@2017](#) [Линк](#)
52. Takács, T., Cseresnyés, I., Kovács, R., Parádi, I., Kelemen, B., Szili-Kovács, T., & Füzy, A. (2018). Symbiotic effectivity of dual and tripartite associations on soybean (*Glycine max* L. Merr.) cultivars inoculated with *Bradyrhizobium japonicum* and AM fungi. *Frontiers in Plant Science*, 9, 1631., [@2018](#) [Линк](#)
53. Georgiev, Y. N., Ognyanov, M. H., & Denev, P. N. (2020). The ancient Thracian endemic plant *Haberlea rhodopensis* Friv. And related species: A review. *Journal of Ethnopharmacology*, 249, 112359., [@2020](#) [Линк](#)
9. Sárvári, E., Mihailova, G., Solti, A., Keresztes, A., Velitchkova, M., Georgieva, K. Comparison of thylakoid structure and organization in sun and shade *Haberlea rhodopensis* populations under desiccation and rehydration. *Journal of Plant Physiology*, 171, 17, Elsevier, 2014, DOI:10.1016/j.jplph.2014.07.015, 1591-1600. SJR:1.004, ISI IF:2.557
- Цитира се е:
54. Charuvi D, Nevo R, Shimoni E, Naveh L, Zia A, Adam Z, ... Reich Z. Photoprotection conferred by changes in photosynthetic protein levels and organization during dehydration of a homoiochlorophyllous resurrection plant, *Plant Physiology*, 167, 4, 1554-1565, [@2015](#) [Линк](#)
55. Hou X, Fu A, Garcia VJ, Buchanan BB, Luan S. PSB27: A thylakoid protein enabling *Arabidopsis* to adapt to changing light intensity, *Proceedings of the National Academy of Sciences*, 112, 5, 1613-1618, [@2015](#) [Линк](#)
56. Mladenov P, Finazzi G, Bligny R, Moyanova D, Zasheva D, Boisson AM, ... Djilianov D. In vivo spectroscopy and NMR metabolite fingerprinting approaches to connect the dynamics of photosynthetic and metabolic phenotypes in resurrection plant *Haberlea rhodopensis* during desiccation and recovery, *Frontiers in plant science*, 6, 564, [@2015](#) [Линк](#)
57. Muneer S, Jeong BR. Genotypic Variation under Fe Deficiency Results in Rapid Changes in Protein Expressions and Genes Involved in Fe Metabolism and Antioxidant Mechanisms in Tomato Seedlings (*Solanum lycopersicum* L.), *International journal of molecular sciences*, 16, 12, 28022-28037, [@2015](#) [Линк](#)
58. Manzoor, H., Athar, H. U. R., Rasul, S., Kanwal, T., Anjam, M. S., Qureshi, M. K., ... & Ashraf, M. (2016). Avenues for improving drought tolerance in crops by ABA regulation. *Water Stress and Crop Plants: A Sustainable Approach*, 177-193., [@2016](#) [Линк](#)
59. Costa, M. C. D., Cooper, K., Hilhorst, H. W., & Farrant, J. M. (2017). Orthodox seeds and resurrection plants: Two of a kind?. *Plant Physiology*, 175(2), 589-599., [@2017](#) [Линк](#)
60. Farrant, J. M., Cooper, K., Dace, H. J. W., Bentley, J., Hilgart, A. (2017) Desiccation Tolerance. In: PLANT STRESS PHYSIOLOGY, 2ND EDITION Pages: 217-252, [@2017](#) [Линк](#)

61. Tan, T., Sun, Y., Luo, S., Zhang, C., Zhou, H., Lin, H. Efficient modulation of photosynthetic apparatus confers 1.000 desiccation tolerance in the resurrection plant *Boea hygrometrica*. *Plant Cell Physiology*, 58(11), 1976-1990, 2017, [@2017](#) [Линк](#)
  62. Garcés, M., Ulloa, M., Miranda, A., & Bravo, L. A. (2018). Physiological and ultrastructural characterisation of a 1.000 desiccation-tolerant filmy fern, *Hymenophyllum caudiculatum*: Influence of translational regulation and ABA on recovery. *Plant Biology*, 20(2), 288-295., [@2018](#) [Линк](#)
  63. Liu, J., Moyankova, D., Djilianov, D., & Deng, X. (2019). Common and Specific Mechanisms of Desiccation 1.000 Tolerance in Two Gesneriaceae Resurrection Plants. Multiomics Evidences. *Frontiers in plant science*, 10, 1067., [@2019](#) [Линк](#)
  64. Wang, Y., Zhang, B., Jiang, D., & Chen, G. (2019). Silicon improves photosynthetic performance by optimizing 1.000 thylakoid membrane protein components in rice under drought stress. *Environmental and experimental botany*, 158, 117-124., [@2019](#) [Линк](#)
  65. Fernández-Marín, B., Nadal, M., Gago, J., Fernie, A. R., López-Pozo, M., Artetxe, U., ... & Verhoeven, A. (2020). 1.000 Born to revive: molecular and physiological mechanisms of double tolerance in a paleotropical and resurrection plant. *New Phytologist*, 226(3), 741-759., [@2020](#) [Линк](#)
  66. Georgiev, Y. N., Ognyanov, M. H., & Denev, P. N. (2020). The ancient Thracian endemic plant *Haberlea rhodopensis* Friv. And related species: A review. *Journal of Ethnopharmacology*, 249, 112359., [@2020](#) [Линк](#)
  67. Jiang, D., Gao, W., & Chen, G. (2023). Toxic effects of lanthanum (III) on photosynthetic performance of rice 1.000 seedlings: Combined chlorophyll fluorescence, chloroplast structure and thylakoid membrane protein assessment. *Ecotoxicology and Environmental Safety*, 267, 115627., [@2023](#) [Линк](#)
  68. Singh, A., Pankaczi, F., Rana, D., May, Z., Tolnai, G., & Fodor, F. (2023). Coated Hematite Nanoparticles 1.000 Alleviate Iron Deficiency in Cucumber in Acidic Nutrient Solution and as Foliar Spray. *Plants*, 12(17), 3104., [@2023](#) [Линк](#)
  69. Fernández-Marín, B., Nadal, M., Perera-Castro, A. V., & García-Plazaola, J. I. (2024). Photosynthetic Strategies 1.000 of Desiccation-Tolerant Organisms from a Functional Perspective. In *Handbook of Photosynthesis* (pp. 659-675). CRC Press., [@2024](#) [Линк](#)
  70. Yu, R., Song, Q., Wang, G., Wu, L., Ruan, J., Yang, C., ... & Wang, J. (2023). Desiccation Tolerance Mechanisms 1.000 of Resurrection Plant *Selaginella pulvinata*. *Environmental and Experimental Botany*, 218, 105602., [@2024](#) [Линк](#)
  
  10. Solti, A., **Mihailova, G.**, Sárvári, E., **Georgieva, K.** Antioxidative defence mechanisms contributes to desiccation tolerance in *Haberlea rhodopensis* population naturally exposed to high irradiation. *Acta Biologica Szegediensis*, 58, 1, 2014, 11-14
- Цитира се е:
71. Mladenov P, Zasheva D, Djilianov D, Tchorbadjieva M. TOWARDS PROTEOMICS OF DESICCATION 1.000 TOLERANCE IN THE RESURRECTION PLANT HABERLEA RHODOPENSI, COMPTES RENDUS DE L'ACADEMIE BULGARE DES SCIENCES, 68, 1, 59-64, [@2015](#) [Линк](#)
  72. Apostolova, E. G., Kokova, V., Peychev, Z., Peycheva, S., & Apostolov, A. (2017). EFFECT OF FUCOIDAN, 1.000 HABERLEA RHODOPENSI AND PROPOLIS ON MOBILIZATION OF THE CD34+ STEM CELLS IN RATS. *FARMACIA*, 65(4), 567-570., [@2017](#) [Линк](#)
  73. Mamik, S., & Sharma, A. D. (2017). Antioxidant response of the invasive alien species *Parthenium hysterophorus* 1.000 L. under abiotic stress conditions with special emphasis on boiling-stable antioxidant enzymes. *Botanica Serbica*, 41 (1), 25-36, [@2017](#) [Линк](#)
  74. Mamik, S., & Sharma, A. D. (2017). Protective role of boiling stable antioxidant enzymes in invasive alien species 1.000 of *Lantana* exposed to natural abiotic stress like conditions. *Russian Journal of Biological Invasions*, 8(1), 75-86., [@2017](#) [Линк](#)
  75. Liu, J., Moyankova, D., Lin, C. T., Mladenov, P., Sun, R. Z., Djilianov, D., & Deng, X. (2018). Transcriptome 1.000 reprogramming during severe dehydration contributes to physiological and metabolic changes in the resurrection plant *Haberlea rhodopensis*. *BMC Plant Biology*, 18(1), 351., [@2018](#) [Линк](#)

---

2015

---

11. Rapparini, F., Neri, L., **Mihailova, G.**, Petkova, S., Georgieva, K. Growth irradiance affects the photoprotective mechanisms of the resurrection angiosperm *Haberlea rhodopensis* Friv. in response to desiccation and rehydration at morphological, physiological and biochemical levels. *Environmental and Experimental Botany*, 113, Elsevier, 2015, DOI:10.1016/j.envexpbot.2015.01.007, 67-79. SJR:1.038, ISI IF:3.359

Цитира се:

76. Choi, H. G., Moon, B. Y., & Kang, N. J. (2016). Correlation between Strawberry (*Fragaria ananassa* Duch.) Productivity and Photosynthesis-Related Parameters under Various Growth Conditions. *Frontiers in Plant Science*, 7, 1607., [@2016 Линк](#)
77. Farrant, J. M., Cooper, K., Dace, H. J. W., Bentley, J., Hilgart, A. (2017) Desiccation Tolerance. In: *PLANT STRESS PHYSIOLOGY*, 2ND EDITION Pages: 217-252, [@2017 Линк](#)
78. Fernández-Marín, B., Neuner, G., Kuprian, E., Laza, J. M., García-Plazaola, J. I., & Verhoeven, A. (2018). First evidence of freezing tolerance in a resurrection plant: insights into molecular mobility and zeaxanthin synthesis in the dark. *Physiologia plantarum*, 163, 472-489., [@2018 Линк](#)
79. Han, Y., Wu, J., Tian, Y., Zha, T., Jia, X., Bourque, C., ... & Zhang, M. (2018). Light Energy Partitioning and Photoprotection in an Exotic Species (*Salix Psammophila*) Grown in a Semi-Arid Area of Northwestern China. *Forests*, 9(6), 341., [@2018 Линк](#)
80. Verhoeven, A., García-Plazaola, J. I., & Fernández-Marín, B. (2018). Shared mechanisms of photoprotection in photosynthetic organisms tolerant to desiccation or to low temperature. *Environmental and Experimental Botany*, 154, 66-79., [@2018 Линк](#)
81. Yu, R.-P., Wang, G.-X., Li, F., (...), Yang, C.-M., Qu, Y.-H. (2019) Photosynthesis response of resurrection plant *Selaginella pulvinata* detached leaves to dehydration and rehydration. *Zhiwu Shengli Xuebao/Plant Physiology Journal*, 55(12), pp. 1806-1816, [@2019 Линк](#)
82. Fernández-Marín, B., Nadal, M., Gago, J., Fernie, A. R., López-Pozo, M., Artetxe, U., ... & Verhoeven, A. (2020). Born to revive: molecular and physiological mechanisms of double tolerance in a paleotropical and resurrection plant. *New Phytologist*, 226(3), 741-759., [@2020 Линк](#)
83. Georgiev, Y. N., Ognyanov, M. H., & Denev, P. N. (2020). The ancient Thracian endemic plant *Haberlea rhodopensis* Friv. And related species: A review. *Journal of Ethnopharmacology*, 249, 112359., [@2020 Линк](#)
84. Nadal, M., Brodrribb, T.J., Fernández-Marín, B., García-Plazaola, J.I., Arzac, M.I., López-Pozo, M., Perera-Castro, A.V., Gulías, J., Flexas, J. and Farrant, J.M., 2021. Differences in biochemical, gas exchange and hydraulic response to water stress in desiccation tolerant and sensitive fronds of the fern *Anemia caffrorum*. *New Phytologist*, [@2021 Линк](#)
85. Nadal, M., Perera-Castro, A. V., Gulías, J., Farrant, J. M., & Flexas, J. (2021). Resurrection plants optimize photosynthesis despite very thick cell walls by means of chloroplast distribution. *Journal of Experimental Botany*, 72(7), 2600-2610., [@2021 Линк](#)
86. Tebele S.M., Marks R.A., Farrant J.M. (2021) Two Decades of Desiccation Biology: A Systematic Review of the Best Studied Angiosperm Resurrection Plants. *Plants*, 10(12), 2784., [@2021 Линк](#)
87. Yu, R., Cheng, Y., Pu, Y., Li, F., & Lu, S. (2021). In Vitro Propagation of Resurrection Plant *Selaginella pulvinata* Using Frond Tips as Explants. *HortScience*, 1(aop), 1-5., [@2021 Линк](#)
88. Nadal, M., Carriquí, M., Badel, E., Cochard, H., Delzon, S., King, A., ... & Torres-Ruiz, J. M. (2023). Photosynthesis, leaf hydraulic conductance and embolism dynamics in the resurrection plant *Barbacenia purpurea*. *Physiologia Plantarum*, 175(5), e14035., [@2023 Линк](#)
89. Qiu, J., Cai, C., Shen, M., Gu, X., Zheng, L., Sun, L., ... & Zou, L. (2023). Responses of growth, yield and fruit quality of strawberry to elevated CO<sub>2</sub>, LED supplemental light, and their combination in autumn through spring greenhouse production. *Plant Growth Regulation* 102(2), 351-365, [@2024 Линк](#)
90. Vieira, E. A., Gaspar, M., Caldeira, C. F., Munné-Bosch, S., & Braga, M. R. (2024). Desiccation tolerance in the resurrection plant *Barbacenia graminifolia* involves changes in redox metabolism and carotenoid oxidation. *Frontiers in Plant Science*, 15, 1344820., [@2024 Линк](#)

---

2016

---

12. Georgieva, K, Mihailova, G. Drought Tolerance of Photosynthesis. Handbook of photosynthesis, Third edition, CRC Press, Taylor & Francis Group, 2016, ISBN:9781482230734, 683-696

Цитира се е:

91. Smolikova, G., Kreslavski, V., Shiroglazova, O., Bilova, T., Sharova, E., Frolov, A., Medvedev, S. Photochemical 1.000 activity changes accompanying the embryogenesis of pea (*Pisum sativum*) with yellow and green cotyledons. Functional Plant Biology, 45(2), 228-235, 2017, [@2017](#) [Линк](#)

---

2017

---

13. Georgieva K, Dagnon S, Gesheva E, Bojilov D, Mihailova G, Doncheva S. Antioxidant defense during desiccation of the resurrection plant *Haberlea rhodopensis*. Plant Physiology and Biochemistry, 114, Elsevier, 2017, ISSN:0981-9428, SCOPUS, DOI:<https://doi.org/10.1016/j.plaphy.2017.02.021>, 51-59. JCR-IF (Web of Science):2.718

Цитира се е:

92. Challabathula D, Zhang Q, Bartels D. Protection of photosynthesis in desiccation-tolerant resurrection plants. 1.000 Journal of Plant Physiology, 227, 84-92, [@2018](#) [Линк](#)
93. Ariff MA, Tukiman S, Razak NA, Osman MS, Jaapar J. Optimization of supercritical fluid extraction of Mariposa 1.000 *Christia Vespretilionis* leaves towards antioxidant using response surface methodology. In: Journal of Physics: Conference Series, 1349, 1, 012054. IOP Publishing., [@2019](#) [Линк](#)
94. Hell, A. F., Gasulla, F., González-Hourcade, M., Del Campo, E. M., Centeno, D. C., & Casano, L. M. (2019). 1.000 Tolerance to cyclic desiccation in lichen microalgae is related to habitat preference and involves specific priming of the antioxidant system. Plant and Cell Physiology, 60(8), 1880-1891., [@2019](#) [Линк](#)
95. Shi J, Wang W, Lin Y, Xu K, Xu Y, Ji D, Chen C, Xie C. Insight into transketolase of *Pyropia haitanensis* under 1.000 desiccation stress based on integrative analysis of omics and transformation. BMC plant biology, 19, 1, 475., [@2019](#) [Линк](#)
96. Xiang T, Xu H, Yan W, Song D. Effects of hispidulin on the activity of human liver cytochrome P450 enzymes. 1.000 Lat Am J Pharm, 38, 1, 57-62., [@2019](#) [Линк](#)
97. Ariff, M. A., Nazri, N. N., Abdul Razak, N. A., Osman, M. S., & Jaapar, J. (2020). Optimization of Supercritical 1.000 Fluid Extraction for Mariposa *Christia Vespretilionis* Leaves Towards Yield by Using Response Surface Methodology. Recent Innovations in Chemical Engineering (Formerly Recent Patents on Chemical Engineering), 13(2), 170-178., [@2020](#) [Линк](#)
98. Bentley, J., Olsen, E. K., Moore, J. P., & Farrant, J. M. (2020). The phenolic profile extracted from the desiccation- 1.000 tolerant medicinal shrub *Myrothamnus flabellifolia* using Natural Deep Eutectic Solvents varies according to the solvation conditions. Phytochemistry, 173, 112323., [@2020](#) [Линк](#)
99. Fernández-Marín, B., Nadal, M., Gago, J., Fernie, A. R., López-Pozo, M., Artetxe, U., ... & Verhoeven, A. (2020). 1.000 Born to revive: molecular and physiological mechanisms of double tolerance in a paleotropical and resurrection plant. New Phytologist, 226(3), 741-759., [@2020](#) [Линк](#)
100. García-Fontana, C., Vilchez, J. I., & Manzanera, M. (2020). Proteome comparison between natural desiccation- 1.000 tolerant plants and drought-protected *Capsicum annuum* plants by *Microbacterium* sp. 3J1. Frontiers in Microbiology, 11, 1537., [@2020](#) [Линк](#)
101. Georgiev, Y. N., Ognyanov, M. H., & Denev, P. N. (2020). The ancient Thracian endemic plant *Haberlea rhodopensis* Friv. And related species: A review. Journal of Ethnopharmacology, 249, 112359., [@2020](#) [Линк](#)
102. Wang, Y., Gao, S., He, X., Li, Y., Zhang, Y., & Chen, W. (2020). Response of total phenols, flavonoids, minerals, 1.000 and amino acids of four edible fern species to four shading treatments. PeerJ, 8, e8354., [@2020](#) [Линк](#)
103. Wen, J., Wang, W., Xu, K., Ji, D., Xu, Y., Chen, C., & Xie, C. (2020). Comparative analysis of proteins involved 1.000 in energy metabolism and protein processing in *Pyropia haitanensis* at different salinity levels. Frontiers in Marine Science, 7, 415, [@2020](#) [Линк](#)

104. Gasulla, F., Del Campo, E. M., Casano, L. M., & Guéra, A. (2021). Advances in Understanding of Desiccation Tolerance of Lichens and Lichen-Forming Algae. *Plants*, 10(4), 807., @2021 [Линк](#)
105. Xiang, T., Xu, H., Yan, W., Song, D. (2021) Effects of hispidulin on the activity of human liver cytochrome p450 enzymes. *Latin American Journal of Pharmacy*, 40(9), 2036-2042, @2021 [Линк](#)
106. Alejo-Jacuinde, G., Kean-Galeno, T., Martínez-Gallardo, N., Tejero-Díez, J. D., Mehltreter, K., Délano-Frier, J. P., ... & Herrera-Estrella, L. (2022). Viability markers for determination of desiccation tolerance and critical stages during dehydration in *Selaginella* species. *Journal of Experimental Botany*, 73(12), 3898-3912., @2022 [Линк](#)
107. Bruñás Gómez, I., Casale, M., Barreno, E., & Catalá, M. (2022). Near-Infrared Metabolomic Fingerprinting Study of Lichen Thalli and Phycobionts in Culture: Aquaphotomics of *Trebouxia lynnæ* Dehydration. *Microorganisms*, 10(12), 2444., @2022 [Линк](#)
108. Godevac, D., Ivanović, S., Simić, K., Andelković, B., Jovanović, Ž., & Rakić, T. (2022). Metabolomics study of the desiccation and recovery process in the resurrection plants *Ramonda serbica* and *R. nathaliae*. *Phytochemical Analysis*, 33(6), 961-970., @2022 [Линк](#)
109. Ignatov, I., Huether, F., Neshev, N., Kiselova-Kaneva, Y., Popova, T. P., Bankova, R., ... & Baiti, S. (2022). Research of Water Molecules Cluster Structuring during *Haberlea rhodopensis* Friv. Hydration. *Plants*, 11(19), 2655., @2022 [Линк](#)
110. Ivanova, A., O' Leary, B., Signorelli, S., Falconet, D., Moyankova, D., Whelan, J., ... & Murcha, M. W. (2022). Mitochondrial activity and biogenesis during resurrection of *Haberlea rhodopensis*. *New Phytologist*, 236(3), 943-957., @2022 [Линк](#)
111. Del Campo, E. M., Gasulla, F., Hell, A. F., González-Hourcade, M., & Casano, L. M. (2023). Comparative Transcriptomic and Proteomic Analyses Provide New Insights into the Tolerance to Cyclic Dehydration in a Lichen Phycobiont. *Microbial Ecology*, 1-15., @2023 [Линк](#)
112. Legardón, A., & García-Plazaola, J. I. (2023). Gesneriads, a Source of Resurrection and Double-Tolerant Species: Proposal of New Desiccation-and Freezing-Tolerant Plants and Their Physiological Adaptations. *Biology*, 12(1), 107., @2023 [Линк](#)
113. Santos Reginaldo, F.P., Rodrigues de Souza, L.F., Sotero Chacon, D., Dias Mariano Santos, M., Melo Torres, T., Bezerra da Silva, I., Araújo Roque, A.D., O. Ricart, C.A., Castro, M.S., Carvalho, P.C. and Fontes, W., 2024. Organ-specific proteomes of *Selaginella convoluta* provide insights into its desiccation tolerance mechanisms. *Plant Biosystems-An International Journal Dealing with all Aspects of Plant Biology*, pp.1-8., @2024 [Линк](#)
114. Yu R, Song Q, Wang G, Wu L, Ruan J, Yang C, Li S, Wang J. Desiccation Tolerance Mechanisms of Resurrection Plant *Selaginella pulvinata*. *Environmental and Experimental Botany*. 218, 105602., @2024 [Линк](#)
14. **Mihailova G**, Abakumov D, Büchel C, Dietzel L, **Georgieva K**. Drought-responsive gene expression in sun and shade plants of *Haberlea rhodopensis* under controlled environment. *Plant Molecular Biology Reporter*, 35, 3, Springer, 2017, ISSN:0735-9640 (Print) 1572-9818 (Online), Web of Science, DOI:10.1007/s11105-017-1025-3, 313-322. ISI IF:2.304  
*Цитира се е:*
115. Munné-Bosch, S., & Vincent, C. (2019). Physiological Mechanisms Underlying Fruit Sunburn. *Critical Reviews in Plant Sciences*, 38(2), 140-157., @2019 [Линк](#)
116. Sebastiani, F., Torre, S., Gori, A., Brunetti, C., Centritto, M., Ferrini, F., & Tattini, M. (2019). Dissecting Adaptation Mechanisms to Contrasting Solar Irradiance in the Mediterranean Shrub *Cistus incanus*. *International journal of molecular sciences*, 20(14), 3599., @2019 [Линк](#)
15. **Georgieva K**, Rapparini F, Bertazza G, **Mihailova G**, Sárvári É, Solti Á, Keresztes Á. Alterations in the sugar metabolism and in the vacuolar system of mesophyll cells contribute to the desiccation tolerance of *Haberlea rhodopensis* ecotypes. *Protoplasma*, 254, 1, Springer, 2017, ISSN:0033-183X; eISSN: 1615-6102, Web of Science, DOI:10.1007/s00709-015-0932-0, 193-201. SJR:0.96, ISI IF:2.651  
*Цитира се е:*
117. Durgud, M., Gupta, S., Ivanov, I., Omidbakhshfard, M. A., Benina, M., Alseekh, S., ... & Toneva, V. (2018). Molecular mechanisms preventing senescence in response to prolonged darkness in a desiccation-tolerant plant. *Plant physiology*, 177(3), 1319-1338., @2018 [Линк](#)

118. Liu, J., Moyankova, D., Lin, C. T., Mladenov, P., Sun, R. Z., Djilianov, D., & Deng, X. (2018). Transcriptome reprogramming during severe dehydration contributes to physiological and metabolic changes in the resurrection plant *Haberlea rhodopensis*. *BMC Plant Biology*, 18(1), 351., @2018 [Линк](#)
119. López-Pozo, M., Fernández-Marín, B., García-Plazaola, J. I., & Ballesteros, D. (2018). Desiccation Tolerance in Ferns: From the Unicellular Spore to the Multi-tissular Sporophyte. In *Current Advances in Fern Research* (pp. 401-426). Springer, Cham., @2018 [Линк](#)
120. Kuroki, S., Tsenkova, R., Moyankova, D., Muncan, J., Morita, H., Atanassova, S., & Djilianov, D. (2019). Water molecular structure underpins extreme desiccation tolerance of the resurrection plant *Haberlea rhodopensis*. *Scientific reports*, 9(1), 3049., @2019 [Линк](#)
121. Ovsyannikov, A. Y., & Koteyeva, N. K. (2020). Seasonal movement of chloroplasts in mesophyll cells of two *Picea* species. *Protoplasma*, 257(1), 183-195., @2020 [Линк](#)
122. Legardón, A., & García-Plazaola, J. I. (2023). Gesneriads, a Source of Resurrection and Double-Tolerant Species: Proposal of New Desiccation-and Freezing-Tolerant Plants and Their Physiological Adaptations. *Biology*, 12(1), 107., @2023 [Линк](#)
123. Wang, Y., Wang, W., Chi, X., Cheng, M., Wang, T., Zhan, X., ... & Li, X. (2023). Analysis and Identification of Genes Associated with the Desiccation Sensitivity of *Panax notoginseng* Seeds. *Plants*, 12(22), 3881., @2023 [Линк](#)
124. Aronsson, H., & Solymosi, K. (2024). Diversification of Plastid Structure and Function in Land Plants. In *Plastids: Methods and Protocols* (pp. 63-88). New York, NY: Springer US., @2024 [Линк](#)

**2018**

16. Mihailova, G, Kocheva, K, Goltsev, V, Kalaji, HM, Georgieva, K. Application of a diffusion model to measure ion leakage of resurrection plant leaves undergoing desiccation. *Plant Physiology and Biochemistry*, 125, Elsevier, 2018, ISSN:0981-9428, Web of Science, DOI:10.1016/j.plaphy.2018.02.008, 185-192. SJR (Scopus):1.159, JCR-IF (Web of Science):3.404

*Цитира се е:*

125. Escalante-Magaña C , Aguilar-Caamal LF, Echevarría-Machado I, Medina-Lara F, Cach LS, Martínez-Estevez M. Contribution of glycine betaine and proline to water deficit tolerance in pepper plants, *HortScience*, 54, 6, 1044–1054., @2019 [Линк](#)
126. Chen P, Jung NU, Giarola V, Bartels D. The dynamic responses of cell walls in resurrection plants during dehydration and rehydration. *Frontiers in Plant Science*, 10, 1698., @2020 [Линк](#)
127. Georgiev YN, Ognyanov MH, Denev PN. The ancient Thracian endemic plant *Haberlea rhodopensis* Friv. And related species: A review. *Journal of ethnopharmacology*, 249, 112359, @2020 [Линк](#)
128. Jokar A, Zare H, Zakerin A, Jahromi AA, The Influence of Photo-selective Netting on Tree Physiology and Fruit Quality of Fig (*Ficus carica* L.) Under Rain-fed Conditions, *International Journal of Fruit Science*, 21, 1, 896-910, @2021 [Линк](#)
129. Sun R-Z, Liu J, Wang Y-Y, Deng X, DNA methylation-mediated modulation of rapid desiccation tolerance acquisition and dehydration stress memory in the resurrection plant *Boea hygrometrica*, *PLoS Genetics*, 17, 4, e1009549, @2021 [Линк](#)
130. Caputo L, Amato G, de Bartolomeis P, De Martino L, Manna F, Nazzaro F, ... & Barba AA. Impact of drying methods on the yield and chemistry of *Origanum vulgare* L. essential oil. *Scientific Reports*, 12(1), 1-11, @2022 [Линк](#)
131. De Martino L, Caputo L, Amato G, Iannone M, Barba AA, De Feo V. Postharvest Microwave Drying of Basil (*Ocimum basilicum* L.): The Influence of Treatments on the Quality of Dried Products. *Foods*, 11(7), 1029, @2022 [Линк](#)
132. Apicella M, Amato G, de Bartolomeis P, Barba AA, De Feo V. 2023. Natural Food Resource Valorization by Microwave Technology: Purslane Stabilization by Dielectric Heating. *Foods*, 12, 23, 4247., @2023 [Линк](#)
133. Legardón A, García-Plazaola JI. Gesneriads, a Source of Resurrection and Double-Tolerant Species: Proposal of New Desiccation-and Freezing-Tolerant Plants and Their Physiological Adaptations. *Biology*, 12(1), 107, @2023 [Линк](#)

134. Shitova AS, Petrenko DB, Okina OI, Vasiliev NV. Fluoride influence on the growth, morphological and biochemical parameters, and elemental composition of *Triticum aestivum* in a model experiment. ProQuest, 56(1), 55-74., @2023
135. Xu X, Yang H, Suo X, Liu H, Jing D, Zhang Y, Dang J, Wu D, He Q, Xia Y, Wang S, Liang G, Guo Q. EjFAD8 Enhances the Low-Temperature Tolerance of Loquat by Desaturation of Sulfoquinovosyl Diacylglycerol (SQDG) International Journal of Molecular Science, 24(8), 6946, @2023 [Линк](#)
136. Zou J, Jiang C, Qiu S, Duan G, Wang G, Li D, Yu S, Zhao D, Sun W. An *Ustilaginoidea virens* glycoside hydrolase 42 protein is an essential virulence factor and elicits plant immunity as a PAMP. Molecular Plant Pathology, 1-16, @2023 [Линк](#)
137. Liu, J., Wang, Y., Chen, X., Tang, L., Yang, Y., Yang, Z., ... & Deng, X. (2024). Specific metabolic and cellular mechanisms of the vegetative desiccation tolerance in resurrection plants for adaptation to extreme dryness. *Planta*, 259(2), 47., @2024 [Линк](#)
138. Su, T.H., Shen, Y., Chiang, Y.Y., Liu, Y.T., You, H.M., Lin, H.C., Kung, K.N., Huang, Y.M. and Lai, C.M., 2024. Species selection as a key factor in the afforestation of coastal salt-affected lands: Insights from pot and field experiments. *Journal of Environmental Management*, 360, 121126., @2024 [Линк](#)
139. Udpuary, S., Ullah, H., Himanshu, S. K., Tisarum, R., Praseartkul, P., Cha-um, S., & Datta, A. (2024). Effects of microbial biofertilizer on growth, physio-biochemical traits, fruit yield, and water productivity of okra under drought stress. *Biocatalysis and Agricultural Biotechnology*, 103125., @2024 [Линк](#)
140. Wang, Y., Shen, Y., Dong, W., Cai, X., Yang, J., Chen, Y., ... & Sun, X. (2024). PHD17 acts as a target of miR1320 to negatively control cold tolerance via JA-activated signaling in rice. *The Crop Journal.*, @2024 [Линк](#)

**2019**

17. Mihailova G, Stoyanova Z, Rodeva R, Bankina B, Bimšteine G, Georgieva K. Physiological changes in winter wheat genotypes in response to the *Zymoseptoria tritici* infection. *Photosynthetica*, 57, 2, Institute of Experimental Botany, ASCR, Prague, 2019, ISSN:1573-9058, Web of Sciences, DOI:10.32615/ps.2019.054, 428-437. JCR-IF (Web of Science):2.562

*Цитира се е:*

141. Aydogdu M. Impact of seedling infection of *Septoria tritici* blotch on bread wheat. *Journal of Animal and Plant Sciences* 31, 2, 500-508, @2021 [Линк](#)
142. Otero-Blanca, A., Pérez-Llano, Y., Reboledo-Blanco, G., Lira-Ruan, V., Padilla-Chacon, D., Folch-Mallol, J.L., Sánchez-Carbente, M.D.R., Ponce De León, I. and Batista-García, R.A., 2021. *Physcomitrium patens* Infection by *Colletotrichum gloeosporioides*: Understanding the Fungal-Bryophyte Interaction by Microscopy, Phenomics and RNA Sequencing. *Journal of Fungi*, 7(8), p.677., @2021 [Линк](#)
143. Ghiasi Noei, F., Imami, M., Didaran, F., (...), Feechan, A., Mirzadi Gohari, A. (2022). Stb6 mediates stomatal immunity, photosynthetic functionality, and the antioxidant system during the *Zymoseptoria tritici*-wheat interaction. *Frontiers in Plant Science*, 13, 1004691., @2022 [Линк](#)
144. Spyroglou, I., Rybka, K., Czembor, P., Piaskowska, D., Pernisová, M., & Matysik, P. (2022). Higher alterations in leaf fluorescence parameters of wheat cultivars predict more extensive necrosis in response to *Zymoseptoria tritici*. *Plant Pathology.*, @2022 [Линк](#)
145. Silva, B. N., Picanço, B. B., Martins, S. C., & Rodrigues, F. A. (2023). Impairment of the photorespiratory pathway on tomato leaves during the infection process of *Septoria lycopersici*. *Physiological and Molecular Plant Pathology*, 125, 102020., @2023 [Линк](#)
18. Dolchinkova V, Andreeva T, Georgieva K, Mihailova G, Balashev K. Desiccation-induced alterations in surface topography of thylakoids from resurrection plant *Haberlea rhodopensis* studied by atomic force microscopy, electrokinetic and optical measurements. *Physiologia Plantarum*, 166, 2, Wiley, 2019, ISSN:0031-9317 (print); 1399-3054 (web), Web of Science, DOI:<https://doi.org/10.1111/ppl.12807>, 585-595. JCR-IF (Web of Science):4.148

*Цитира се е:*

146. Li, W. Q., Qing, T., Li, C. C., Li, F., Ge, F., Fei, J. J., & Peijnenburg, W. J. (2020). Integration of subcellular partitioning and chemical forms to understand silver nanoparticles toxicity to lettuce (*Lactuca sativa* L.) under different exposure pathways. *Chemosphere*, 127349, @2020 [Линк](#)

147. Alejo-Jacuinde, G., & Herrera-Estrella, L. (2022). Exploring the High Variability of Vegetative Desiccation Tolerance in Pteridophytes. *Plants*, 11(9), 1222., @2022 [Линк](#)

## 2020

19. Georgieva K, Mihailova G, Velitchkova M, Popova AV. Recovery of photosynthetic activity of resurrection plant *Haberlea rhodopensis* from drought- and freezing-induced desiccation. *Photosynthetica*, 58, 4, Institute of Experimental Botany, Academy of Sciences of the Czech Republic, 2020, ISSN:1573-9058, DOI:10.32615/ps.2020.044, 911-921. JCR-IF (Web of Science):3.189

Цитира се е:

148. Fernández-Marín, B., Arzac, M.I., López-Pozo, M., Laza, J.M., Roach, T., Stegner, M., Neuner, G. and García- Plazaola, J.I., 2021. Frozen in the dark: interplay of night-time activity of xanthophyll cycle, xylem attributes, and desiccation tolerance in fern resistance to winter. *Journal of Experimental Botany*, 72(8), pp.3168-3184., @2021 [Линк](#)
149. Holmlund, H.I., 2021. Synergistic adaptations: freezing tolerance is associated with desiccation tolerance and activation of violaxanthin de-epoxidase in wintergreen ferns. *Journal of Experimental Botany*, 72(8), pp.2814-2817., @2021 [Линк](#)
150. Soorni, J., Kazemitarab, S.K., Kahrizi, D. et al. Genetic analysis of freezing tolerance in camelina [Camelina sativa (L.) Crantz] by diallel cross of winter and spring biotypes. *Planta* 253, 9 (2021), @2021 [Линк](#)
151. Živković, S., Skorić, M., Ristić, M., Filipović, B., Milutinović, M., Perišić, M. and Puač, N., 2021. Rehydration Process in Rustyback Fern (*Aspleniumceterach* L.): Profiling of Volatile Organic Compounds. *Biology*, 10(7), p.574, @2021 [Линк](#)
152. Huang, H. X., Cao, Y., Xin, K. J., Liang, R. H., Chen, Y. T., & Qi, J. J. (2022). Morphological and physiological changes in *Artemisia selengensis* under drought and after rehydration recovery. *Frontiers in Plant Science*, 13, 851942., @2022 [Линк](#)
153. Ivanova, A., O' Leary, B., Signorelli, S., Falconet, D., Moyankova, D., Whelan, J., ... & Murcha, M. W. (2022). Mitochondrial activity and biogenesis during resurrection of *Haberlea rhodopensis*. *New Phytologist*, 236(3), 943-957., @2022 [Линк](#)
154. Szalai, G., Dernovics, M., Gondor, O. K., Tajti, J., Molnár, A. B., Lejmel, M. A., ... & Janda, T. (2022). Mutations in Rht-B1 Locus May Negatively Affect Frost Tolerance in Bread Wheat. *International Journal of Molecular Sciences*, 23(14), 7969., @2022 [Линк](#)
155. Legardón, A., & García-Plazaola, J. I. (2023). Gesneriads, a Source of Resurrection and Double-Tolerant Species: Proposal of New Desiccation-and Freezing-Tolerant Plants and Their Physiological Adaptations. *Biology*, 12(1), 107., @2023 [Линк](#)
156. Ferioun, M., Bouhraoua, S., Belahcen, D., Zouitane, I., Srhiouar, N., Louahlia, S., & El Ghachoui, N. (2024). PGPR Consortia Enhance Growth and Yield in Barley Cultivars Subjected to Severe Drought Stress and Subsequent Recovery. *Rhizosphere*, 100926., @2024 [Линк](#)
157. Fernández-Marín, B., Nadal, M., Perera-Castro, A. V., & García-Plazaola, J. I. (2024). Photosynthetic Strategies of Desiccation-Tolerant Organisms from a Functional Perspective. In *Handbook of Photosynthesis* (pp. 659-675). CRC Press., @2024 [Линк](#)

20. Mihailova G, Solti Á, Sárvári É, Keresztes Á, Rapparini F, Velitchkova M, Simova-Stoilova L, Aleksandrov V, Georgieva K. Freezing tolerance of photosynthetic apparatus in the homoiochlorophylloous resurrection plant *Haberlea rhodopensis*. *Environmental and Experimental Botany*, 178, 104157, Elsevier, 2020, ISSN:0098-8472, DOI:<https://doi.org/10.1016/j.envexpbot.2020.104157>, JCR-IF (Web of Science):5.545

Цитира се е:

158. Tebele S.M., Marks R.A., Farrant J.M. (2021) Two Decades of Desiccation Biology: A Systematic Review of the Best Studied Angiosperm Resurrection Plants. *Plants*, 10(12), 2784., @2021 [Линк](#)
159. Alejo-Jacuinde, G., Herrera-Estrella, L. Exploring the High Variability of Vegetative Desiccation Tolerance in Pteridophytes. *Plants* 11(9), 1222, @2022 [Линк](#)

160. Liu, X., Quan, W., Bartels, D. Stress memory responses and seed priming correlate with drought tolerance in plants: an overview. *Planta* 255(2), 45, @2022 [Линк](#)
161. Legardón, A., & García-Plazaola, J. I. (2023). Gesneriads, a Source of Resurrection and Double-Tolerant Species: Proposal of New Desiccation-and Freezing-Tolerant Plants and Their Physiological Adaptations. *Biology*, 12(1), 107., @2023 [Линк](#)
162. Aronsson, H., & Solymosi, K. (2024). Diversification of Plastid Structure and Function in Land Plants. In *Plastids: Methods and Protocols* (pp. 63-88). New York, NY: Springer US., @2024 [Линк](#)
163. Fernández-Marín, B., Nadal, M., Perera-Castro, A. V., & García-Plazaola, J. I. (2024). Photosynthetic Strategies of Desiccation-Tolerant Organisms from a Functional Perspective. In *Handbook of Photosynthesis* (pp. 659-675). CRC Press., @2024 [Линк](#)
164. Gupta, S., Petrov, V., Garg, V., Mueller-Roeber, B., Fernie, A. R., Nikoloski, Z., & Gechev, T. (2024). The genome of Haberlea rhodopensis provides insights into the mechanisms for tolerance to multiple extreme environments. *Cellular and Molecular Life Sciences*, 81(1), 117., @2024 [Линк](#)
165. Liu, J., Wang, Y., Chen, X., Tang, L., Yang, Y., Yang, Z., ... & Deng, X. (2024). Specific metabolic and cellular mechanisms of the vegetative desiccation tolerance in resurrection plants for adaptation to extreme dryness. *Planta*, 259(2), 47., @2024 [Линк](#)
166. Song, X., & Gao, T. (2024). Experimental investigation of freeze injury temperatures in trees and their contributing factors based on electrical impedance spectroscopy. *Frontiers in Plant Science*, 15, 1326038., @2024 [Линк](#)

**2021**

21. Georgieva K, Mihailova G, Gigova L, Dagnon S, Simova-Stoilova L, Velitchkova M. The role of antioxidant defense in freezing tolerance of resurrection plant Haberlea rhodopensis. *Physiology and Molecular Biology of Plants*, 27, 5, Springer Nature, 2021, ISSN:0971-5894, DOI:<https://doi.org/10.1007/s12298-021-00998-0>, 1119-1133. SJR (Scopus):0.75, JCR-IF (Web of Science):2.391

Цитира се е:

167. Alejo-Jacuinde, G., & Herrera-Estrella, L. (2022). Exploring the High Variability of Vegetative Desiccation Tolerance in Pteridophytes. *Plants*, 11(9), 1222., @2022 [Линк](#)
168. Brúňáková K, Bálintová M, Petříková L, Čellárová E. 2022. Does phenotyping of Hypericum secondary metabolism reveal a tolerance to biotic/abiotic stressors?. *Frontiers in Plant Science*, 13: 1042375. SJR: 1.36; Q1., @2022 [Линк](#)
169. Ghitti E, Rolli E, Crotti E, Borin S. 2022. Flavonoids Are Intra-and Inter-Kingdom Modulator Signals. *Microorganisms*, 10(12):2479. SJR: 0.86; Q2, @2022 [Линк](#)
170. Liu, X., Quan, W., & Bartels, D. (2022). Stress memory responses and seed priming correlate with drought tolerance in plants: An overview. *Planta*, 255(2), 1-14., @2022 [Линк](#)
171. Legardón, A., & García-Plazaola, J. I. (2023). Gesneriads, a Source of Resurrection and Double-Tolerant Species: Proposal of New Desiccation-and Freezing-Tolerant Plants and Their Physiological Adaptations. *Biology*, 12(1), 107., @2023 [Линк](#)
172. Zhang Z, Gu Y, Mao Q, Wang J. 2023. "Physiological response to low temperature of four genotypes of Cyclocarya paliurus and their preliminary evaluation to cold resistance." *Forests*, 14(8):1680. SJR: 0.65; Q1., @2023 [Линк](#)

**2022**

22. Georgieva K, Mihailova G, Fernández-Marín B, Bertazza G, Govoni A, Irati Arzac M, Laza JM, Vilas JL, García-Plazaola JL, Rapparini F. Protective Strategies of Haberlea rhodopensis for Acquisition of Freezing Tolerance: Interaction between Dehydration and Low Temperature. *International Journal of Molecular Sciences*, 23, 23, MDPI, 2022, DOI:<https://doi.org/10.3390/ijms232315050>, 15050. JCR-IF (Web of Science):6.208

Цитира се е:

173. Fan, J., Zhang, Y., Sun, H., Duan, R., Jiang, Y., Wang, X., ... Yu, H. (2024). Overexpression of soybean GmDHN9 gene enhances drought resistance of transgenic *Arabidopsis*. *GM Crops & Food*, 15(1), 118–129., @2024 [Линк](#)
23. Popova AV, Borisova P, **Mihailova G, Georgieva K.** Antioxidative response of *Arabidopsis thaliana* to combined action of low temperature and high light illumination when lutein is missing. *Acta Physiologae Plantarum*, 44, 10, Springer, 2022, DOI:<https://doi.org/10.1007/s11738-021-03342-x>, 1-15. JCR-IF (Web of Science):2.354
- Цитира се е:
174. Wu, Y., Cai, X., & Tang, Y. (2022). Outcomes of Low-Temperature Stress on Biological Alterations within *Pothos Epipremnum aureum* Leaves. *Life*, 12(9), 1432., @2022 [Линк](#)
24. **Mihailova G, Vasileva I, Gigova L, Gesheva E, Simova-Stoilova L, Georgieva K.** Antioxidant Defense during Recovery of Resurrection Plant *Haberlea rhodopensis* from Drought- and Freezing-Induced Desiccation. *Plants*, 11, 2, MDPI, 2022, DOI:<https://doi.org/10.3390/plants11020175>, 175. JCR-IF (Web of Science):4.658
- Цитира се е:
175. Alejo-Jacuinde G, Herrera-Estrella L. Exploring the High Variability of Vegetative Desiccation Tolerance in *Pteridophytes*. *Plants*, 11(9), 1222., @2022 [Линк](#)
176. Mladenov P, Zasheva D, Planchon S, Leclercq CC, Falconet D, Moyet L, Brugiére S, Moyankova D, Tchorbadjieva M, Ferro M, Rolland N, Renaut J, Djilianov D, Deng X. 2022. Proteomics Evidence of a Systemic Response to Desiccation in the Resurrection Plant *Haberlea rhodopensis*. *International Journal of Molecular Sciences*, 23(15): 8520. SJR: 1.18; Q1., @2022 [Линк](#)
177. Bhatt, U., & Soni, V. (2023). Study of biochemical and biophysical adjustments during transition from desiccation-to-fully-hydrated states in *Riccia gangetica* and *Semibarbula orientalis*. *Vegetos*, 36(2), 550-558., @2023 [Линк](#)
178. Bittencourt, P. P., Alves, A. F., Ferreira, M. B., da Silva Irineu, L. E. S., Pinto, V. B., & Olivares, F. L. (2023). Mechanisms and Applications of Bacterial Inoculants in Plant Drought Stress Tolerance. *Microorganisms*, 11(2), 502., @2023 [Линк](#)
179. Legardón A, García-Plazaola JI. Gesneriads, a Source of Resurrection and Double-Tolerant Species: Proposal of New Desiccation-and Freezing-Tolerant Plants and Their Physiological Adaptations. *Biology*, 12(1), 107., @2023 [Линк](#)
180. Fernández-Marín, B., Nadal, M., Perera-Castro, A. V., & García-Plazaola, J. I. (2024). Photosynthetic Strategies of Desiccation-Tolerant Organisms from a Functional Perspective. In *Handbook of Photosynthesis* (pp. 659-675). CRC Press., @2024 [Линк](#)
181. Liu, J., Wang, Y., Chen, X., Tang, L., Yang, Y., Yang, Z., ... & Deng, X. (2024). Specific metabolic and cellular mechanisms of the vegetative desiccation tolerance in resurrection plants for adaptation to extreme dryness. *Planta*, 259(2), 47., @2024 [Линк](#)
25. Popova AV, Vladkova R, Borisova P, **Georgieva K, Mihailova G, Velikova V, Tsonev T**, Ivanov AG. Photosynthetic response of lutein-deficient mutant *lut2* of *Arabidopsis thaliana* to low temperature at high light. *Photosynthetica*, 60, 1, Institute of Experimental Botany, Academy of Sciences of the Czech Republic, 2022, ISSN:0300-3604, DOI:[10.32615/ps.2022.009](https://doi.org/10.32615/ps.2022.009), 110-120. JCR-IF (Web of Science):2.482
- Цитира се е:
182. PRÁŠIL, O., KAŇA, R., & Govindjee, G. (2022). Special issue in honor of Prof. George C. Papageorgiou. *PHOTOSYNTHETICA*, 60(1), 136-146., @2022 [Линк](#)
183. Yagoubi A, Mahjoubi Y, Giannakis S, Rzigi T, Djebali W, Chouari R. The silver lining of antibiotic resistance: Bacterial-mediated reduction of tetracycline plant stress via antibiotropy. *Plant Physiology and Biochemistry* v. 204, art. 108093, @2023 [Линк](#)
26. **Georgieva K, Popova AV, Mihailova G**, Ivanov AG, Velitchkova M. Limiting steps and the contribution of alternative electron flow pathways in the recovery of the photosynthetic functions after freezing-induced desiccation of *Haberlea rhodopensis*. *Photosynthetica*, 60, 1, Institute of Experimental Botany, Academy of Sciences of the Czech Republic, 2022, ISSN:0300-3604, DOI:[10.32615/ps.2022.008](https://doi.org/10.32615/ps.2022.008), 136-146. JCR-IF (Web of Science):2.482
- Цитира се е:

184. PRÁŠIL, O., KAŇA, R., & Govindjee, G. (2022). Special issue in honor of Prof. George C. Papageorgiou. 1.000 PHOTOSYNTHETICA, 60(1), 136-146., @2022 [Линк](#)
185. Legardón, A., & García-Plazaola, J. I. (2023). Gesneriads, a Source of Resurrection and Double-Tolerant 1.000 Species: Proposal of New Desiccation-and Freezing-Tolerant Plants and Their Physiological Adaptations. Biology, 12(1), 107., @2023 [Линк](#)
27. Mihailova G, Christov NK, Sárvári É, Solti Á, Hembrom R, Solymosi K, Keresztes Á, Velitchkova M, Popova AV, Simova-Stoilova L, Todorovska E, Georgieva K. Reactivation of the Photosynthetic Apparatus of Resurrection Plant Haberlea rhodopensis during the Early Phase of Recovery from Drought- and Freezing-Induced Desiccation. Plants, 11, 17, MDPI, 2022, ISSN:2223-7747, DOI:10.3390/plants11172185, 2185. JCR-IF (Web of Science):4.658

Цитира се е:

186. Legardón A, García-Plazaola JI. (2023). Gesneriads, a Source of Resurrection and Double-Tolerant Species: 1.000 Proposal of New Desiccation-and Freezing-Tolerant Plants and Their Physiological Adaptations. Biology, 12(1), 107., @2023 [Линк](#)
187. Aronsson, H., Solymosi, K. Diversification of Plastid Structure and Function in Land Plants. Methods in Molecular 1.000 Biology 2776, pp. 63-88, @2024 [Линк](#)
188. Basavaraj, P.S., Jangid, K.K., Babar, R., Rane, J., Boraiah, K.M., Harisha, C.B., Halli, H.M., Pradhan, A. and 1.000 Reddy, K.S., 2024. Genetic variation in deficit moisture stress tolerance of Cicer accessions revealed by chlorophyll fluorescence. Genetic Resources and Crop Evolution, pp.1-15., @2024 [Линк](#)
189. Fernández-Marín, B., Nadal, M., Perera-Castro, A. V., & García-Plazaola, J. I. (2024). Photosynthetic Strategies 1.000 of Desiccation-Tolerant Organisms from a Functional Perspective. In Handbook of Photosynthesis (pp. 659-675). CRC Press., @2024 [Линк](#)

## 2023

28. Mihailova G, Gashi B, Krastev N, Georgieva K. Acquisition of Freezing Tolerance of Resurrection Species from Gesneriaceae, a Comparative Study. Plants, 12, 9, MDPI, 2023, DOI:<https://doi.org/10.3390/plants12091893>, 1893. JCR-IF (Web of Science):4.5

Цитира се е:

190. Liu, J., Wang, Y., Chen, X., Tang, L., Yang, Y., Yang, Z., ... & Deng, X. (2024). Specific metabolic and cellular 1.000 mechanisms of the vegetative desiccation tolerance in resurrection plants for adaptation to extreme dryness. Planta, 259(2), 47., @2024 [Линк](#)

29. Mihailova G, Solti Á, Sárvári É, Hunyadi-Gulyás É, Georgieva K. Protein Changes in Shade and Sun Haberlea rhodopensis Leaves during Dehydration at Optimal and Low Temperatures. Plants, 12, 2, MDPI, 2023, DOI:<https://doi.org/10.3390/plants12020401>, 401. JCR-IF (Web of Science):4.5

Цитира се е:

191. Djiljanov, D., Moyankova, D., Mladenov, P., Topouzova-Hristova, T., Kostadinova, A., Staneva, G., ... & Simova-Stoilova, L. (2024). Resurrection Plants—A Valuable Source of Natural Bioactive Compounds: From Word-of-Mouth to Scientifically Proven Sustainable Use. Metabolites, 14(2), 113., @2024 [Линк](#)

30. Mihailova G, Tchorbadjieva M, Rakleova G, Georgieva K. Differential Accumulation of sHSPs Isoforms during Desiccation of the Resurrection Plant Haberlea rhodopensis Friv. under Optimal and High Temperature. Life, 13, 1, MDPI, 2023, DOI:<https://doi.org/10.3390/life13010238>, 238. JCR-IF (Web of Science):3.2

Цитира се е:

192. Gomathi, R., Kohila, S., Viswanathan, R., Krishnapriya, V., Appunu, C., Kumar, R.A., Alagupalamuthirsolai, M., 1.000 Manimekalai, R., Elayaraja, K. and Kaverinathan, K., 2024. Comparative Proteomic Analysis of High-Temperature Response in Sugarcane (*Saccharum* spp.). Sugar Tech, pp.1-15., @2024 [Линк](#)

31. Popova A, **Mihailova G, Geneva M, Peeva V, Kirova E, Sichanova M, Dobrikova A, Georgieva K.** Different Responses to Water Deficit of Two Common Winter Wheat Varieties: Physiological and Biochemical Characteristics. *Plants*, 12, 12, MDPI, 2023, ISSN:2223-7747, DOI:<https://doi.org/10.3390/plants12122239>, 2239. JCR-IF (Web of Science):4.5

Цитира се е:

193. Rathore R, Mishra M, Pareek A, Singla-Pareek S, Concurrent improvement of rice grain yield and abiotic stress tolerance by overexpression of cytokinin activating enzyme LONELY GUY (OsLOG), *Plant Physiol and Biochem*, 211, 108635, **@2024** [Линк](#)

194. Vedenicheva, N., & Kosakivska, I. (2024). The Regulatory Role of Cytokinins in Adaptive Responses of Cereal Plants. In: *Plant Science Research and Practices*, 83-110, **@2024** [Линк](#)

195. Zhang, B., Li, C., Yi, G., Liu, R. (2024) Physiological, biochemical and morphological responses of *Haloxylon ammodendron* and *Calligonum caput-medusae* to drought stress. *Arid Zone Pesearch*, 41(7), 1177–1184, **@2024** [Линк](#)

32. **Georgieva K, Mihailova G, Gigova L**, Popova AV, Velitchkova M, **Simova-Stoilova L**, Sági-KazárM, Zelenyánszki H, Solymosi K, Solti Á. Antioxidative Defense, Suppressed Nitric Oxide Accumulation, and Synthesis of Protective Proteins in Roots and Leaves Contribute to the Desiccation Tolerance of the Resurrection Plant *Haberlea rhodopensis*. *Plants*, 12, 15, MDPI, 2023, DOI:<https://doi.org/10.3390/plants12152834>, 2834. JCR-IF (Web of Science):4.5

Цитира се е:

196. Allagulova, C., Avalbaev, A., Lubyanova, A., Plotnikov, A., Yuldashev, R., & Lastochkina, O. (2023). Nitric Oxide (NO) Improves Wheat Growth under Dehydration Conditions by Regulating Phytohormone Levels and Induction of the Expression of the TADHN Dehydrin Gene. *Plants*, 12(23), 4051., **@2023** [Линк](#)



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